

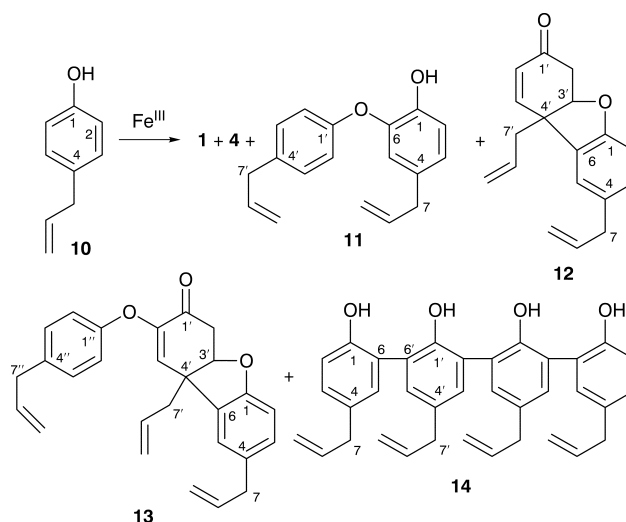
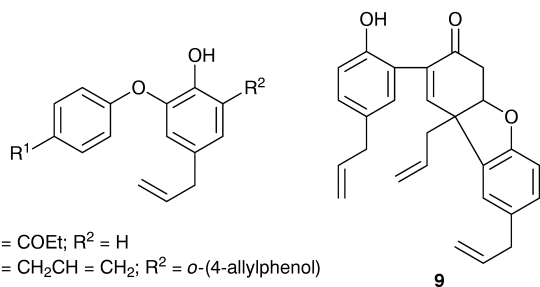
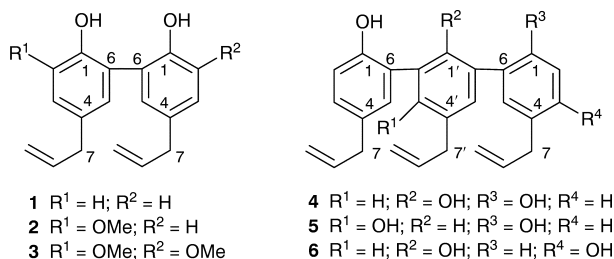
Biomimetic Synthesis of *Illicium* Oligomeric Neolignans†

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Six products have been isolated from the oxidative coupling of 4-allylphenol in the presence of Fe^{III}; the distribution of products suggests that oligomeric neolignans associated with the family *Illicium* may also be non-enzymic products of oxidative coupling.

The genus *Illicium* is characterised by a variety of oligomeric neolignans which are linked through the aromatic ring (C₆): these include *ortho,ortho* (*o,o*)-linked dimers^{1,2} [magnolol **1**; 2,2'-dihydroxy-3-methoxy-5,5'-di(prop-2-enyl) biphenyl **2** and dehydrodieugenol **3**], *o,o*-linked trimers²⁻⁴ (dunnianol **4**; macrathanol **5** and simonsinol **6**), dimers and trimers involving *o,O*-linkage (isomagnolone⁴ **7** and isodunnianol^{1,2} **8**) and a *o,o*-/*o,p*-linked trimer **9**.⁵ [A few neolignans have also been reported which are coupled through the phenylpropanoid 'tail' (C₃).]⁶



Scheme 1

coupled product magnolol **1** (15%), the trimeric *o,o*-coupled product dunnianol **4** (2%) and the *o,O*-coupled product isomagnolone **11** (1%). Use of K₃Fe(CN)₆¹⁶ in place of FeCl₃ gave a better overall yield (44%) but also resulted in a qualitatively more complex mixture of dimeric, trimeric and tetrameric *o,o*-, *o,O*- and *o,p*-linked products (**1**, **4**, **11-14**; Scheme 1), in which **4** and **12** were the major components. All compounds were characterized by NMR (compounds **12-14** are reported for the first time) and ¹H and ¹³C resonances were rigorously assigned using 2D NMR techniques (e.g. HSQC, HMBC, ¹H-¹H COSY), see Experimental section and Table 1.

Gottlieb has proposed that oxidative coupling reactions might account for the biosynthesis of a wide range of neolignans⁷ although there has been relatively little experimental work to validate this hypothesis.⁸ Consequently, we decided to subject 4-allylphenol **10**, a putative biogenetic precursor to the *Illicium* neolignans, to oxidative coupling *in vitro* in order to compare the distribution of products formed with that reported *in vivo*. The oxidative coupling of phenols is a well established process⁹ and Ho *et al.* have previously described a photochemical oxidative coupling of 4-allylphenol which yielded magnolol **1**¹⁰ (magnolol has also been prepared by coupling of aromatic halides in the presence of Pd⁰).¹¹ Anodic oxidations of 2-methoxy-4-allylphenol and 2,6-dimethoxy-4-allylphenol are reported to yield complex mixtures of dimeric products.^{12,13} We herein report the results of the oxidative coupling of 4-allylphenol in the presence of Fe^{III}.

4-Allylphenol **10**, required for oxidative coupling studies, was obtained in 46% yield by treatment of commercially available 4-allylanisole with boron tribromide.¹⁴ Treatment of **10** with FeCl₃¹⁵ produced the dimeric *o,o*-

The distribution of oxidatively coupled products obtained from 4-allylphenol *in vitro*, in which it is possible to identify dimeric, trimeric and tetrameric structures formed as a result of *o,o*-, *o,p*- and *o,O*-coupling reactions, bears a striking resemblance to that reported for natural products from the genus *Illicium*, with the exception that no tetrameric neolignans have as yet been described from Nature (interestingly, however, no oligomers higher than the tetramer were isolated from *in vitro* oxidation). This similarity in chemistry, including the co-occurrence of compounds **1** and **4**, suggests that an analogous non-enzymic oxidative coupling process, operating on 4-allylphenol *in vivo*, may be sufficient to account for the formation of neolignans from the *Illiciaceae*.

Experimental

General Methods.—Chemical shifts are expressed in ppm (δ) relative to TMS as int. standard. All NMR experiments were run on a Bruker DRX 500 instrument. Two dimensional spectra were recorded with 1024 data points in F₂ and 256 data points in F₁. MS were recorded in EI mode at 70 eV on a Finnigan-MAT 95 MS spectrometer. IR spectra were recorded in CHCl₃ on a BIO-RAD FT S-7 IR spectrometer. Column chromatography was performed using silica gel 60–200 μm (Merck).

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Table 1 ^{13}C NMR data for compounds **1**, **4**, **10** and **11–14**

Atom ^a	Compound						
	1	4	10	11	12	13^b	14
1 (C)	151.1	151.4	153.5	145.7	157.0	156.9	151.2
2 (CH)	116.7	117.2	115.5	115.9	110.1	110.3	117.0
3 (CH)	131.2	130.0	129.7	124.6	129.4	129.5	129.8
4 (C)	133.1	133.3	132.1	132.6	131.0	131.5	133.1
5 (CH)	129.9	131.6	129.7	118.9	122.9	122.7	131.5
6 (C)	123.8	126.0	115.5 (CH)	143.4	133.2	133.5	124.8
7 (CH ₂)	39.4	39.7	39.2	39.4	39.6	39.4	39.4
8 (CH ₂)	137.5	137.7	137.8	137.5	137.5	137.6	137.6
9 (CH ₂)	115.8	116.0	115.3	115.7	115.7	115.8	115.8
1' (C)		147.9		155.1	195.2	189.7	147.8
2' (CH ₂)		125.0 (C)		117.9 (CH)	38.7	39.9	126.1 (C)
3' (CH)		131.8		129.9	84.6	84.2	131.7
4' (C)		134.0		135.2	48.4	49.1	133.9
5' (CH)		131.8		129.9	148.4	128.3	131.7
6' (CH)		125.0		117.9	127.1	147.6 (C)	126.0 (C)
7' (CH ₂)		39.5		39.4	40.5	41.2	39.4
8' (CH)		137.4		137.4	132.1	132.2	137.2
9' (CH ₂)		116.3		115.9	119.6	119.8	116.1

^aMultiplicity established from DEPT. ^bAdditional peaks for compound **13**: 154.4 (C), C-1"; 117.9 (CH), C-2"; 129.5 (CH), C-3"; 135.1 (C), C-4"; 129.5 (CH), C-5"; 117.9 (CH), C-6"; 39.7 (CH₂), C-7"; 137.4 (CH), C-8"; 115.7 (CH₂), C-9".

Oxidative Coupling of 10 in the Presence of FeCl₃.—A solution of **10** in 95% EtOH (2.5 g, 10 ml) was added dropwise to an aqueous solution of FeCl₃ (7.03 g, 625 ml). A constant stream of air was maintained through the stirred solution for 48 h and the mixture was then acidified and extracted with Et₂O. The Et₂O layer was in turn extracted with NaOH (2 M); following acidification of the alkaline extract, a further Et₂O extract was collected which was washed, dried, concentrated under reduced pressure and subjected to gradient column chromatography to yield unreacted **10** (1.45 g), **1** (323 mg), **4** (68 mg) and **11** (29 mg).

Oxidative Coupling of 10 in the Presence of K₃Fe(CN)₆.—A K₃Fe(CN)₆ solution (1.85 g, 6.4 ml, 0.88 M) was added dropwise over 30 min to a stirred solution of **10** (0.5 g) in aqueous Na₂CO₃ (0.8 g, 18.7 ml) at 0 °C. The reaction mixture was stirred for 3 h, then acidified and extracted with Et₂O, worked up as before and subjected to gradient column chromatography to yield unreacted **10** (107 mg), **1** (59 mg, 8%), **4** (132 mg, 11%), **11** (35 mg, 5%), **12** (94 mg, 12%), **13** (54 mg, 5%) and **14** (55 mg, 4%).

Magnolol (1).—Oil. IR (CHCl₃) $\nu_{\text{max}}/\text{cm}^{-1}$: 3549, 3267 (br), 3082, 3011, 2907, 1639, 1497, 1215. HREIMS m/z (intensity %): 266.1305 (100) (M⁺, C₁₈H₁₈O₂ requires 266.1307), 248 (5), 247 (9), 225 (15). δ_{H} (CDCl₃): 7.12 (2 H, dd, $J = 8.2, 2.1$ Hz, H-3), 7.08 (2 H, d, $J = 2.1$ Hz, H-5), 6.94 (2 H, d, $J = 8.2$ Hz, H-2), 5.95 (2 H, m, H-8), 5.05 (4 H, m, H-9), 3.35 (4 H, d, $J = 6.7$ Hz, H-7).

Dunnianol (4).—Solid. Mp 133.5–134.5 °C. IR (CHCl₃) $\nu_{\text{max}}/\text{cm}^{-1}$: 3545, 3315 (br), 3085, 3011, 2926, 1640, 1506, 1215. HREIMS m/z (intensity %): 398.1882 (75) (M⁺, C₂₇H₂₆O₃ requires 398.1882); 316 (13), 287 (100), 266 (29). δ_{H} (CDCl₃): 7.14 (2 H, s, H-3'/5'), 7.12 (2 H, d, $J = 2.1$ Hz, H-5), 7.10 (2 H, dd, $J = 8.1, 2.1$ Hz, H-3), 6.92 (2 H, d, $J = 8.1$ Hz, H-2), 5.96 (3 H, m, H-8/8'), 5.10 (6 H, m, H-9/9'), 3.36 (6 H, d, $J = 6.1$ Hz, H-7/7').

4-Allylphenol (10).—Oil. δ_{H} (CDCl₃): 7.14 (1 H, br, s, 1-OH), 7.02 (2 H, d, $J = 8.3$ Hz, H-3/5), 6.77 (2 H, d, $J = 8.3$ Hz, H-2/6), 5.95 (1 H, m, H-8), 5.05 (2 H, m, H-9), 3.31 (2 H, d, $J = 6.7$ Hz, H-7).

Isomagnolol (11).—Oil IR (CHCl₃) $\nu_{\text{max}}/\text{cm}^{-1}$: 3566, 3011, 2978, 2922, 1636, 1506, 1229. HREIMS m/z (intensity %): 266.1306 (100) (M⁺, C₁₈H₁₈O₂ requires 266.1307), 175 (8), 134 (12), 133 (12). δ_{H} (CDCl₃): 7.15 (2 H, d, $J = 8.4$ Hz, H-3'/5'), 6.96 (1 H, d, $J = 8.2$ Hz, H-2), 6.93 (2 H, d, $J = 8.4$ Hz, H-5), 7.10 (2 H, dd, $J = 8.1, 2.1$ Hz, H-3), 1.8 Hz, H-3), 6.70 (1 H, d, $J = 1.8$ Hz, H-5), 5.95 (2 H, m, H-8/8'), 5.49 (1 H, s, 1-OH), 5.05 (4 H, m, H-9/9'), 3.36 (2 H, d, $J = 6.6$ Hz, H-7), 3.24 (2 H, d, $J = 6.6$ Hz, H-7).

Compound 12.—Oil. IR (CHCl₃) $\nu_{\text{max}}/\text{cm}^{-1}$: 3013, 2905, 1686, 1639, 1487, 1250. HREIMS m/z (intensity %): 266.1305 (91) (M⁺, C₁₈H₁₈O₂ requires 266.1307), 239 (13), 225 (100), 197 (16), 184 (14). δ_{H} (CDCl₃): 7.02 (1 H, d, $J = 1.7$ Hz, H-5), 6.99 (1 H, dd, $J = 8.2, 1.7$ Hz, H-3), 6.72 (1 H, d, $J = 8.2$ Hz, H-2), 6.49 (1 H, dd, $J = 10.2, 1.8$ Hz, H-5'), 5.99 (1 H, d, $J = 10.2$ Hz, H-6'), 5.94 (1 H, m, H-8), 5.78 (1 H, m, H-8'), 5.19 (2 H, m, H-9'), 5.07 (2 H, m, H-9), 4.80 (1 H, m, H-3'), 3.33 (2 H, d, $J = 6.7$ Hz, H-7), 3.00 (1 H, dd, $J = 17.6, 2.8$ Hz, H-2'), 2.79 (1 H, dd, $J = 14.2, 6.7$ Hz, H-7'), 2.74 (1 H, dd, $J = 17.6, 4.2$ Hz, H-2'), 2.66 (1 H, dd, $J = 14.2, 8.1$ Hz, H-7).

Compound 13.—Oil. IR (CHCl₃) $\nu_{\text{max}}/\text{cm}^{-1}$: 3078, 3011, 2914, 1697, 1643, 1607, 1502, 1225. HREIMS m/z (intensity %): 398.1882 (100) (M⁺, C₂₇H₂₆O₃ requires 398.1882), 357 (8). δ_{H} (CDCl₃): 7.05 (2 H, d, $J = 8.4$ Hz, H-3'/5'), 7.00 (1 H, d, $J = 8.1$, H-3), 6.92 (1 H, s, H-5), 6.78 (1 H, d, $J = 8.1$ Hz, H-2), 6.73 (2 H, d, $J = 8.4$ Hz, H-2'/6'), 5.95 (2 H, m, H-8/8'), 5.85 (1 H, m, H-8'), 5.81 (1 H, s, H-5'), 5.22 (2 H, m, H-9'), 5.04 (4 H, m, H-9/9'), 4.76 (1 H, m, H-3'), 3.31 (4 H, d, $J = 6.6, 6.7/7/7'$), 3.17 (1 H, dd, $J = 17.6, 2.9$ Hz, H-2'), 2.92 (1 H, dd, $J = 17.6, 5.0$ Hz, H-2'), 2.78 (1 H, dd, $J = 14.2, 6.9$ Hz, H-7'), 2.63 (1 H, dd, $J = 14.2, 8.0$ Hz, H-7').

Compound 14.—Pink solid. mp 168 °C. IR (CHCl₃) $\nu_{\text{max}}/\text{cm}^{-1}$: 3528, 3275 (br), 3082, 3011, 2914, 1636, 1497, 1229. HREIMS m/z (intensity %): 530.2457 (100) (M⁺, C₃₆H₃₄O₄ requires 530.2457). δ_{H} (CDCl₃): 7.20 (2 H, dd, $J = 2.2$ Hz, H-3'), 7.17 (2 H, d, $J = 2.2$ Hz, H-5'), 7.12 (2 H, d, $J = 2.2$ Hz, H-5), 7.09 (2 H, dd, $J = 8.2, 2.2$ Hz, H-3), 6.89 (2 H, d, $J = 8.2$ Hz, H-2), 5.95 (4 H, m, H-8/8'), 5.07 (8 H, m, H-9/9'), 3.41 (2 H, d, $J = 6.7$ Hz, H-7'), 3.36 (4 H, d, $J = 6.7$ Hz, H-7).

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